

Differential induction of muscle atrophy pathways in two mouse models of spinal muscular atrophy

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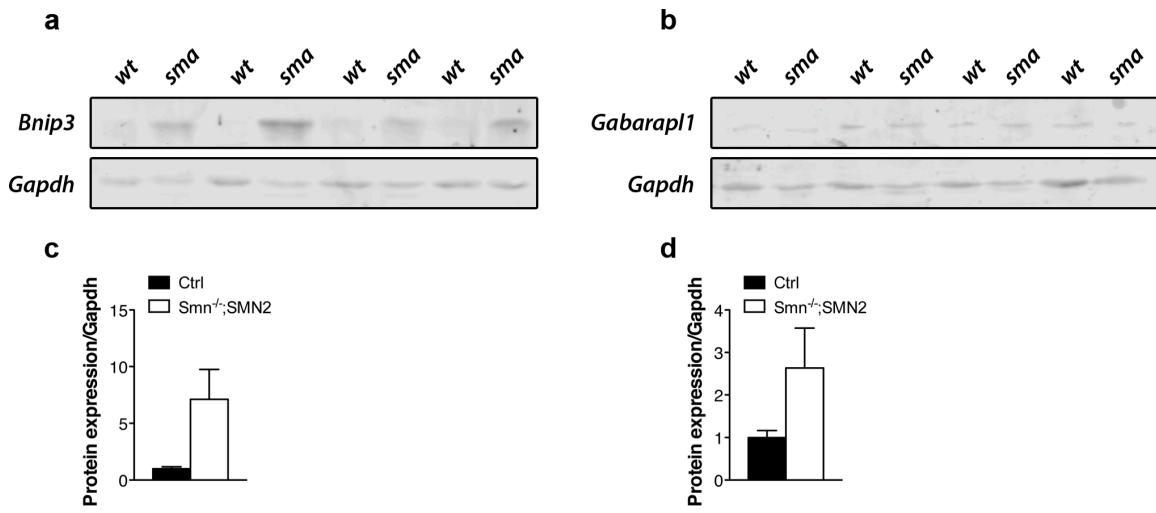
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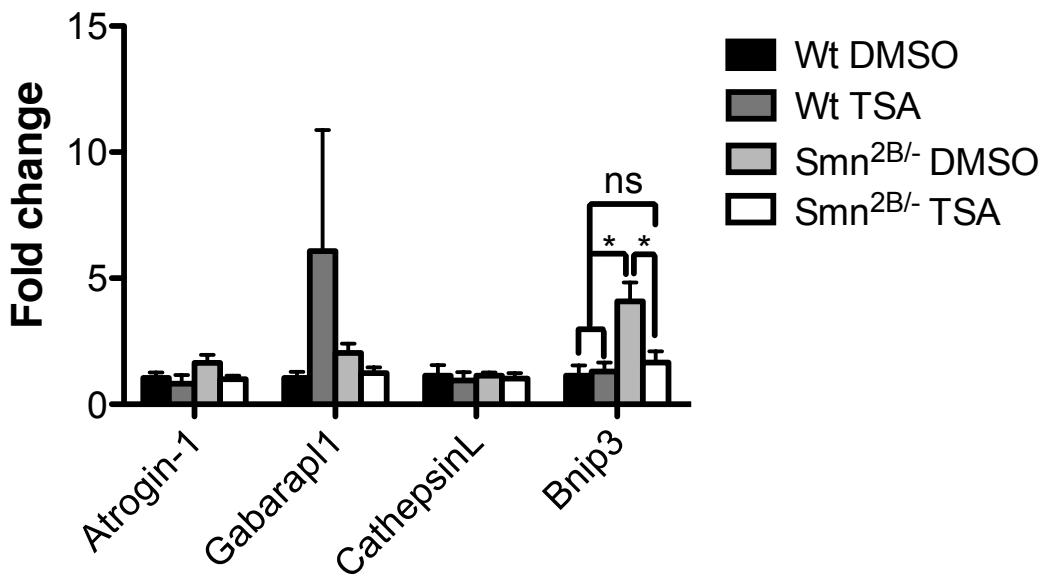
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Supplementary Table S1: List of primers used in this study and their respective annealing temperatures.

Gene target	Forward primer	Reverse primer	T_a
Atrogin-1	CGTCTCACTTCCCCCTCAAG	GACTCCCAGCCATCCAATTAG	57
MuRF1	AGTGTCCATGTCTGGAGGTCGTT	ACTGGAGCACTCCTGCTTAGAT	60
GabarapL1	CATCGTGGAGAAGGGCTCCTA	ATACAGCTGGCCCATGGTAG	62
Bnip3	TTCCACTAGCACCTCTGATGA	GAACACCGCATTACAGAACAA	60
CathepsinL	GTGGACTGTTCTCACGCTCAAG	TCCGTCCCTCGCTTCATAGG	60
FoxO1	CAAAGTACACATAACGCCAATCC	CGTAACTTGATTGCTGTCCTGAA	60
FoxO3	CCTCATCTCAAAGCTGGGTAC	GGTTTCTCTGTAGGTCTCCG	63
FoxO4	AAATGCCAGCCTGGCCAGC	GGCATTCTCCCAGTAACAG	60
Gapdh	TCGGTGTGAACGGATTG	GGTCTCGCTCCTGGAAGA	62



Supplementary Figure S1. *Smn*^{-/-};SMN2 hearts show a trend toward increased Bnip3 and Gabarapl1 protein levels at P5. (a) Western blot of Bnip3. (b) Western blot of Gabarapl1. (c) Quantification of Bnip3 protein expression showing a trend toward elevated protein levels compared to control. (d) Quantification of Gabarapl1 protein expression showing a trend toward elevated protein levels compared to control.



Supplementary Figure S2. TSA administration to *Smn*^{2B/-} mice restores Bnip3 expression in hearts to normal levels. Atrogin-1, Gararapl1 and CathepsinL were unchanged as observed above. TSA treatment did not alter their expression. The increased expression of Bnip3 was effectively brought back to wild type control levels. (N=3 for all experiments; p ≤ 0.05 for *)